

EphA2 chimeric antigen receptor-modified T cells for the immunotherapy of esophageal squamous cell carcinoma

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Background: It is urgent to explore an effective potential therapeutic strategy for ESCC. In recent years, cell-based cancer immunotherapy has become a potentially close for carcinoma therapy. Chimeric antigen receptor (CAR) T cell technology is a kind of adoptive cell therapy technique which has been developed rapidly. We sought to obtain EphA2.CAR-T cell and revealed the ability of EphA2.CAR-T cells to kill esophageal squamous cell carcinoma (ESCC) cells *in vitro*.

Methods: Firstly, the expression and location of EphA2 in ESCC tissues and cells was tested by immunohistochemistry staining and Western blot. Secondly, the second generation of EphA2.CAR was constructed via molecular biology technology, and transduced into T cells to obtain the EphA2.CAR-T cell. The transduction efficacies were assessed using flow cytometry (FCM). Thirdly, the effect of cell killing of EphA2.CAR-T cell on ESCC cells *in vitro* was detected by co-culture experiments. The productions of cytokines (TNF- α and IFN- γ) by EphA2.CAR-T cell after co-culture with ESCC cells were analyzed by ELISA assay.

Results: The expression of EphA2 was significantly upregulated in ESCC tissues and cells ($P<0.05$). EphA2 was expressed on the membrane of ESCC cells, so it could be served as tumor-associated surface antigens (TAA) of CAR for ESCC treatment. The EphA2.CAR-T cell was obtained successfully, and its' transduction efficacies was 61.4% by FCM. The ability of cell killing of EphA2.CAR-T cell was better than that of T cells ($P<0.01$), and demonstrated a dose-dependent cell killing. The results of ELISA assay showed that the levels of TNF- α and IFN- γ in EphA2.CAR-T cells were notably raised compared with T cells ($P<0.05$).

Conclusions: We firstly constructed the second generation of EphA2.CAR and established EphA2.CAR-T cells. The EphA2.CAR-T cells showed a dose-dependent cell killing of ESCC cells, and promoted the production of cytokines *in vitro*. These findings open a new way for treatment of ESCC by immunotherapy in the future.

Keywords: Esophageal squamous cell carcinoma (ESCC); chimeric antigen receptor (CAR); erythropoietin-producing hepatocellular receptor A2 (EphA2); cell killing; TNF- α ; IFN- γ

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Introduction

Esophageal cancer is a common digestive tract tumor and a malignant tumor formed by abnormal hyperplasia of esophageal squamous epithelium or glandular epithelium (1,2). Because 80% of esophageal cancer is composed of esophageal squamous cell carcinoma (ESCC), and based on the improvement of pathological diagnosis and treatment of ESCC, the slight improvement of clinical treatment of ESCC has been promoted. However, the newer and more effective therapies strategies are needed to optimize and promote the clinical treatment of ESCC (3).

Cancer immunotherapy is to kill tumors and inhibit tumor growth by increasing the immunogenicity of tumor cells and the cytotoxicity of effector cells, enhancing immune response for anti-tumor cooperating with the immune system (4-6). T lymphocytes, as the core immune cells that mediate adaptive cellular immune response, could kill tumor cells. In recent years, cell-based cancer immunotherapy has become a potentially close for carcinoma treatment. System modified T cells to express chimeric antigen receptor (CAR), known as CAR T cells, evolve the ordinary t cells to distinguish specific antigen and target tumor cells (7,8). Therefore CARs modified T cell therapy is one of cell-based cancer immunotherapy (9,10). CAR-modified T (CAR-T) cells are not only major histocompatibility complex (MHC)-restricted but also can be engineered to specifically recognize tumor-associated surface antigens (TAA) (11). CAR is a fusion structure formed by a simulation of physiological function of T cell receptor (TCR) (12). Specifically recognition and following signaling cascade is due to the structural characteristics of CAR (13,14). According to the combination of stimulating signal molecules, CAR-T cells were separated into three generations (15). The 2nd generation CAR-T cells were concerned with the introduction of one co-stimulatory molecule (16-18). The 3rd generation CAR-T cells were added another co-stimulatory molecule on the basis of the 2nd generation CAR (19). In order to obtain the best efficacy and the clinical application in the treatment of esophageal cancer in the future, we chose the 2nd generation of the CAR modified T cells to study for tumor cell killing.

The key to treat ESCC with CAR-T cell immunotherapies is to prevent immune escape via targeting antigen on the membrane of carcinoma cells. EphA2 (erythropoietin-producing hepatocellular receptor A2), which is one of the Eph family, becomes known

as a surface antigen (20). EphA2 overexpression brings about carcinogenic effects including oncogenesis, EMT transformation, vasculogenesis and cell growth (21,22). The unique dual role of EphA2 in tumorigenesis and angiogenesis makes it one of the most attractive targets for tumor therapy (23). Recent study of EphA2.CAR-T cells immunotherapy indicated that they were able to recognize and clear out EphA2-positive glioma cells *in vitro*. By the way, leading to glioblastoma multiforme (GBM) tumor regression *in vivo* (20). In ESCC, it was reported that EphA2 overexpression was positive in 40 of the 80 patients (50%) (24). Due to EphA2 is overexpressed in ESCC and EphA2 overexpression correlates with poor prognosis in ESCC (24) and EphA2 is a membrane antigen, a CAR targeting EphA2 could be the ideal solution for the immunotherapy of ESCC.

CAR-T immunotherapy for ESCC has not been reported so far. In the research, we devoted to construct a CAR specifically targeting EphA2 and transduct into T cell, and tested its cell killing efficacy *in vitro*. The research will open a new way for solid tumor treatment of ESCC by the second generation of EphA2.CAR-T cell immunotherapy in the future.

Methods

Tissue specimens

In this research, patients with ESCC were recruited in the Affiliated Hospital of Nantong University from 2010 to 2016. All diagnoses were on the basis of histopathological evidence. All patients were not received preoperative treatments for cancer, such as radiotherapy, chemotherapy or immunotherapy. Some fresh tissues (ESCC tissues and adjacent tissues) after surgical separation were immediately washed with sterile physiological saline before stored at -80 °C. Some tissues were fixed with 4% paraformaldehyde (PFA). The research was authorized by the Ethics Committee of the Affiliated Hospital of Nantong University (No. 2015-070). All patients agreed to use their tissues for scientific research.

Immunohistochemistry

The fixed specimens were dehydrated with graded alcohol, embedded with paraffin and cut into 5-μm-thick sections. The sections were implemented in sodium citrate solution (pH=6.0) and cooked at 100 °C for 10 min for antigen

retrieval. Subsequently, the sections were placed in 3% H₂O₂ for 30 min to eliminate endogenous peroxidases. The sections were reacted with mouse anti-human EphA2 primary antibody (1:200) (Santa Cruz Biotechnology, Dallas, TX) at 4 °C overnight. Then the Pierce streptavidin poly-Horseradish peroxidase (ThermoFisher, Waltham, MA) was used to detect. The next day, some tissues were further incubated with rabbit anti-mouse IgG-TRITC secondary antibody (Millipore, Billerica, MA, USA) for 2 h at room temperature. The staining images were observed under a fluorescence microscope (Zeiss, Oberkochen, Germany).

Cell culture

ESCC cells ECA109 and TE-1 (Jennio Biotech Co.Ltd, Guangzhou, China) were cultured in DMEM media (Gibco, Carlsbad, CA) plus with 10% FCS (fatal cattle serum) (lonsera, Shanghai, China) and 100 U/mL penicillin-streptomycin mixture (Solarbio, Beijing, China). EphA2.CAR-T cells and T cells were maintained in GT-T551 (Takara, Dalian, China) media plus with 10% FCS, 100 U/mL penicillin-streptomycin mixture and 500 IU IL-2 (Novoprotein Scientific, Summit, NJ).

Cell immunofluorescent staining

Cells were cultured on glass coverslips (24-well plates) and fixed with 4% PFA for 20 min at 15–25 °C. Then the glass coverslips were washed 30 min with 0.01% PBS, and incubated with blocking solution containing 10% bovine serum albumin (BSA) for 2 h at 15–25 °C. After that, cells were reacted with mouse anti-human EphA2 primary antibody (1:200) (Santa Cruz Biotechnology, Dallas, TX) at 4 °C overnight. The next day, coverslips were allowed to incubate with rabbit anti-mouse IgG-TRITC secondary antibody (1:200) (Millipore, Billerica, MA) for 2 h at 15–25 °C. Finally, the cells were stained with Hoechst 33258 (Beyotime Institute of Biotechnology, Haimen, China) for 10 min and mounted with anti-fade solution, followed by examination under a fluorescence microscope (Zeiss, Oberkochen, Germany).

Blood donor sample

For all experiments, blood samples were collected with informed consent from healthy volunteers using protocol approved ethically by the Committee of Affiliated

Hospital of Nantong University. Based on operation declaration, lymphoprep density gradient centrifugation kit (STEMCELL Technologies, Vancouver, Canada) was adopted to isolate Peripheral Blood Mononuclear Cells (PBMCs) from blood.

Construction of the second generation EphA2.CAR

The EphA2-specific scFv was derived from the EphA2 monoclonal antibody (mAb) 4H5. GeneArt synthesized the codon-optimized gene, which was cloned into a lentivirus backbone containing the human CD8α hinge and transmembrane domain, 4-1BB co-stimulatory domain and the CD3ζ-chain via 5' Xba I and 3' Bam H I sites. Another CAR containing the irrelevant scFv (FMC63) was served as control. Sequencing was used to verify the cloning of the EphA2.CAR and control.

Transduction of T cells and expansion

Lentiviral supernatant was generated from 293T cells transfected with Lv-CAR-EphA2-GFP, pMD2G and pSPAX2. Human PBMCs which gained from healthy volunteer donors, were stimulated by CD28 antibodies (1 ng/mL) and OKT3 (1 ng/mL) (Novoprotein Scientific, Summit, NJ), and then cultured in media plus with 10% FCS, 100 U/mL penicillin-streptomycin mixture (Beyotime Institute of Biotechnology, Haimen, China) and IL-2 (500 IU/mL). T cells stimulated by OKT3/CD28 were then transduced with the constructed lentiviral vector which encoding EphA2.CAR on RetroNectin coated plates. Meanwhile, non-transduced T cells were used as negative control, and they were activated in the same culture condition. Transduced cells were cultured with IL-15 (5 ng/mL) and IL-7 (100 ng/mL) for 14 to 21 days culture before subsequent analysis. The media and cytokines were changed every 3 days during culture or when passaging the T cells for splitting for expansion. In all the trials, we also compared the function of transduced and non-transduced T cells which obtained from the same donor.

Flow cytometry (FCM)

EphA2.CAR-T cells and none transduced T cells were collected and re-suspended by PBS plus 2% FCS. CAR-T cells transduction efficacies were assessed using FCM by detection of GFP autofluorescence. Data were analyzed by FlowJo 9.3.2 software.

Western blot

Total proteins were isolated from ESCC tissues and cells by using RIPA buffer (Beyotime Institute of Biotechnology, Haimen, China), and quantified by BCA protein assay kit (Promega, Madison, WI, USA). Equivalent amount of proteins were transferred to PVDF membranes (Millipore, Billerica, MA, USA) after separated by SDS-PAGE. After blocking using 5% nonfat milk for 1 h at room temperature, membranes were reacted with mouse anti-human EphA2 primary antibody (1:500) (Santa Cruz Biotechnology, Dallas, TX, USA) in TBST plus with 5% nonfat milk at 4 °C overnight. After reaction with primary antibody, membranes were reacted with secondary HRP-conjugated goat anti-mouse antibody (1:1,000) (Santa Cruz Biotechnology, Dallas, TX, USA) for 2 h at room temperature. After washing, protein bands were measured by using an ECL assay kit (Pierce Protein Biology, Waltham, MA, USA) and imaged by chemiluminescence detection system (Tanon, Shanghai, China). The relative expression of EphA2 was determined as the ratio of gray scale value of EphA2 to β-actin. All experiments were repeated 3 times.

Co-culture assays

To examine the effect of T cells and EphA2.CAR-T cells on ESCC cells killing *in vitro*, target cells (ECA109 and TE-1 cells) were seeded in triplicate in 96-well plate at a concentration of 1×10^4 cells per well. The both target cells were cultured together with T cells or EphA2.CAR-T cells at a serial of E: T (effector to target) ratio (1:1, 2:1, 5:1 and 10:1) for 24 h at 37 °C and 5% CO₂. The morphology of the cells was observed under an inverted microscope (TS100, NIKON Instruments Inc, Japan) and photographed after co-culture.

Crystal violet staining assay

In order to measure the tumor cell killing effects of T cells and EphA2.CAR-T cells, crystal violet staining assay was used. ECA109 and TE-1 cells were seeded triplicate in 96-well plate at a concentration of 1×10^4 cells per well, and co-cultured with T cells or EphA2.CAR-T cells at a serial of E: T ratio (1:1, 2:1, 5:1 and 10:1) for 24 h at 37 °C and 5% CO₂. The supernatant of the culture medium were discarded, and each well was washed 2 times by 0.01% PBS. Cells were fixed with 4% PFA for 15 min, and after that washed with 0.01% PBS for 3 times. Finally cells in

each well were stained by crystal violet staining solution (Solarbio, Beijing, China). The images were observed under an inverted microscope (TS100, NIKON Instruments Inc, Japan) and photographed after staining.

LDH cytotoxicity assay

To assess the cytotoxic activity of EphA2.CAR-T cells, LDH release assay was used. Briefly, ECA109 and TE-1 cells were seeded triplicate in 96-well plate at a concentration of 1×10^4 cells per well, and co-cultured with T cells or EphA2.CAR-T cells at a serial of E: T ratio (1:1, 2:1, 5:1 and 10:1) for 24 h at 37 °C and 5% CO₂. The supernatants were collected after co-culture to analyze the production of LDH by using of a LDH release assay kit (Beyotime Institute of Biotechnology, Haimen, China) according to the operation declaration. OD (optical density) values in each well were read at 490 nm using the multimode reader (Thermo Electron, Waltham, MA). The LDH release activity was analyzed by the following formula: $(OD_{sample} - OD_{blank}) / (OD_{maximum\ release} - OD_{blank}) \times 100\%$.

Analysis of cytokine production

To measure the production of cytokines *in vitro*, ECA109 and TE-1 cells were also seeded triplicate in 96-well plate at a concentration of 1×10^4 cells per well, and co-cultured with T cells or EphA2.CAR-T cells at E: T ratio (5:1) for 24 h at 37 °C and 5% CO₂. Supernatants were collected after co-culture and the levels of TNF-α and IFN-γ were analyzed by ELISA kit (Jiancheng, Nanjing City, China) follow the instructions. The OD values were tested at 450 nm by the multimode reader (Thermo Electron, Waltham, MA, USA).

Statistical analysis

The quantitative date was showed as means ± standard deviation (SD). Student's *t*-test was used to analyze the difference. P<0.05 was regarded as the level of significant in all of the analyses.

Results

EphA2 is expressed in ESCC

Initially, we detected EphA2 expression in ESCC tissues and adjacent tissues by using immunofluorescence and immunohistochemistry staining. It was observed obviously

that the expression of EphA2 on the membrane of ESCC cell and its expression were increased in ESCC tissues, compared with adjacent tissues. It was showed in *Figure 1A,B*. Secondly, we detected EphA2 expression in ESCC cell line ECA109 and TE-1 by cell immunofluorescence. It was also observed obviously that the expression of EphA2 on the membrane of ECA109 and TE-1 cells. It was showed in *Figure 1C*. Thirdly, we detected the expression of EphA2 in ESCC tissues and cells by Western blot. As anticipated, the expressions of EphA2 were significantly increased in ESCC tissues and ESCC cells, compared to adjacent tissues (*Figure 1D*) ($P<0.05$). It was showed in *Figure 1D*. These results confirmed that EphA2 expression was significantly upregulated in ESCC tissues and cells, and EphA2 could be served as TAA of CAR for ESCC treatment.

Construction of the second generation EphA2.CAR T cells

The schematic diagrams of construction of EphA2.CAR and CD19.CAR were showed in *Figure 2A*. CAR-T cells transduction efficacies were assessed using FCM by detection of GFP autofluorescence. The results of FCM analysis showed that the positive percentages of GFP in EphA2.CAR group and CD19.CAR group were 57.6% and 61.4%. It was showed in *Figure 2B*. Compared to the activated T group, the positive percentages of GFP in CAR group were significantly higher.

EphA2.CAR-T cells caused ESCC cell killing in vitro

Firstly, co-culture experiment was used to illustrate the effect of cell killing of EphA2.CAR-T cells on ECA109 and TE-1 cells *in vitro*. The T cells and EphA2.CAR-T cells were observed dose-dependent cell killing after co-culture with target ESCC ECA109 and TE-1 cells at E: T ratio of 1:1 to 10:1 for 24 h under an inverted microscope. When the E: T ratio was 1:1, EphA2.CAR-T cells could eliminate part of ECA109 and TE-1 cells. It was amazing that almost all ECA109 and TE-1 cells were cleared when the E: T ratio reached to 5:1. The ability of cell killing of EphA2.CAR-T cells on ECA109 and TE-1 cells was better than that of T cells. It was showed in *Figure 3A*.

Secondly, crystalline violet staining assay was used to confirm the efficacy of cell killing of EphA2.CAR-T cells on ESCC cells. The results were similar to the observed results under an inverted microscope. It was also showed that the T cells and EphA2.CAR-T cells both demonstrated the dose-dependent cell killing. It was showed in *Figure 3B*.

Thirdly, the cytotoxic activity of EphA2.CAR-T cells for ESCC cells was analyzed by LDH release assay kit. As anticipated, the LDH release in EphA2.CAR-T cells group was significantly increased, compared to the non-transduced T cells group ($P<0.01$). And, the effect of EphA2.CAR-T cells on the LDH release from target ESCC cells ECA109 and TE-1 was also dose-dependent. It was shown in *Figure 3C*. The above results revealed that the EphA2.CAR-T cells had good effects on the target ESCC cells killing.

EphA2.CAR-T cells induce cytokines release

Cytokines are small molecule peptides, secreted by immune cells, could regulate the cell function (such as cell division and differentiation, immune function and inflammation, etc.). So, we tested the production of cytokines by EphA2.CAR-T cells after co-culture with ESCC cells by ELISA. The results of ELISA assay showed that the levels of TNF- α and IFN- γ in EphA2.CAR-T cells were notably raised, compared with non-transduced T cells ($P<0.05$). The secreted TNF- α in EphA2.CAR-T cells after co-culture with ECA109 and TE-1 cell was increased over 2-fold than T cells ($P<0.01$). It was showed in *Figure 4A*. The secreted IFN- γ in EphA2.CAR-T cells after co-culture with ECA109 was increased over 3-fold ($P<0.01$), however IFN- γ was increased only over 2-fold after co-culture with TE-1 cells ($P<0.05$). It was showed in *Figure 4B*. These results indicated that EphA2.CAR-T cells effectively promoted the release of TNF- α and IFN- γ .

Discussion

The development of CAR T technology has brought good news to the treatment of cancer patients, special for leukemia (25). In 2017, a CAR-T cell therapy, Kymriah (TM) (CTL019) by Novartis, for B-cell acute lymphatic leukemia (ALL) was ratified by US food & drug administration (FDA) (26). It was the first FDA approval for CAR-T cell treatment and encouraging researchers to develop various CAR-T cells for immunotherapy in the future. In this study, we had first successfully constructed the second generation EphA2.CAR and established the EphA2.CAR-T cells. We indicated that EphA2.CAR-T cells could kill ESCC lines ECA109 and TE-1 effectively, and promote significantly the release of cytokines.

Tumor immunotherapy is a treatment method for the prevention and treatment of malignant tumors by

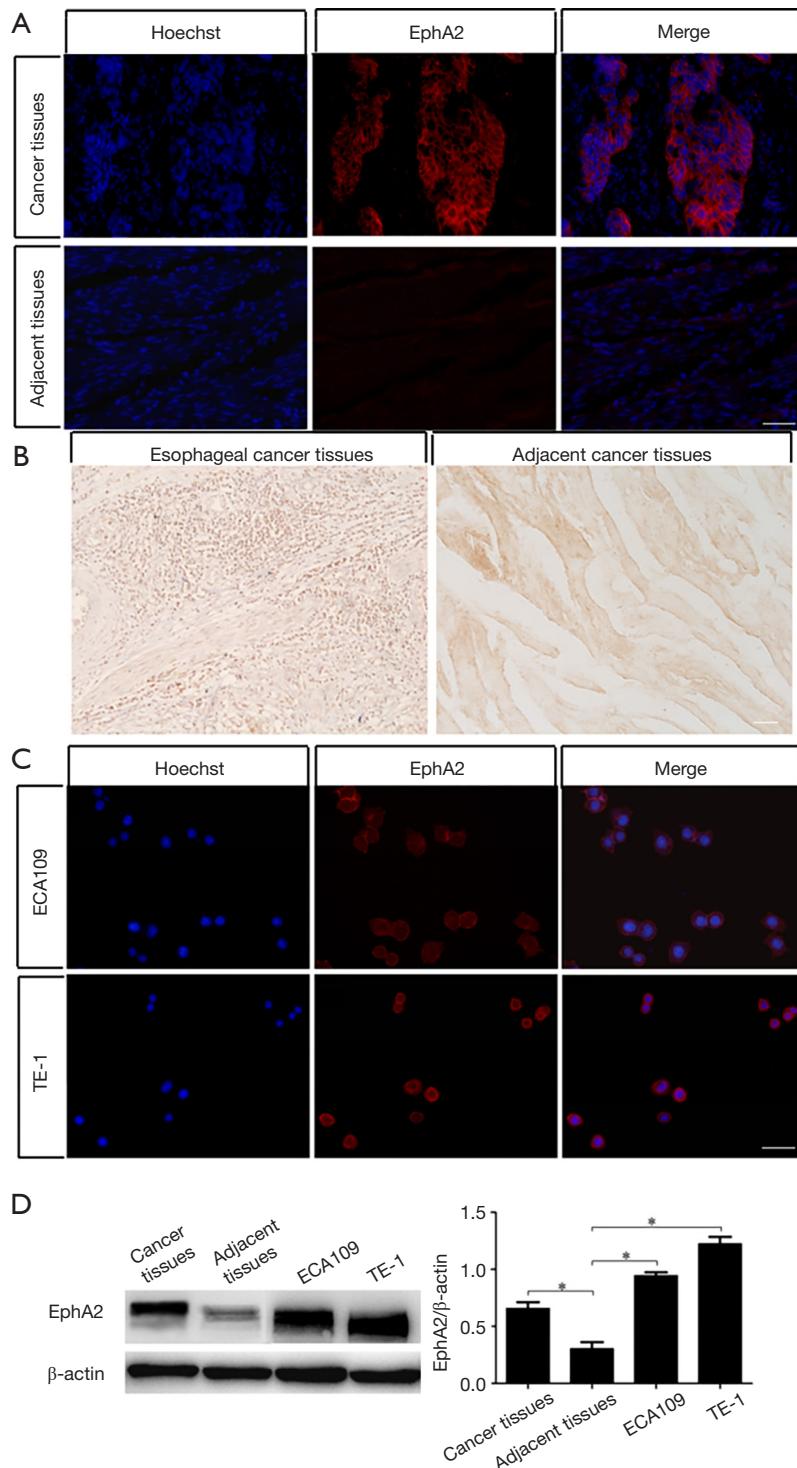


Figure 1 The expression of EphA2 in ESCC tissues and cells. (A) Cellular immunofluorescence staining of EphA2 in ESCC cancer tissues and adjacent tissues; (B) immunohistochemistry staining analysis EphA2 in ESCC cancer tissues and adjacent tissues; (C) cellular immunofluorescence staining of EphA2 in ESCC cell lines ECA109 and TE-1; (D) western blot analysis of EphA2 in ESCC cancer tissues, adjacent tissues, ECA109 and TE-1 cells. Bar =50 μ m. *, $P < 0.05$.

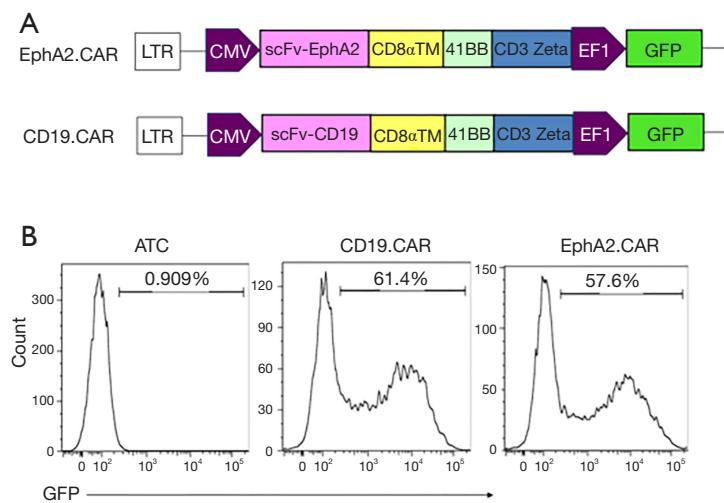


Figure 2 The construction of the second generation EphA2.CAR-T cells. (A) The EphA2.CAR was generated by cloning a single chain variable fragment derived from the EphA2 monoclonal antibody a CD8 α hinge and transmembrane domain, a 4-1BB co-stimulatory domain and a CD3Zeta activation domain into a Lentiviral backbone; (B) EphA2.CAR expression was measured by fluorescence activated cell sorting analysis. ATC, activated T cells; TM, transmembrane region.

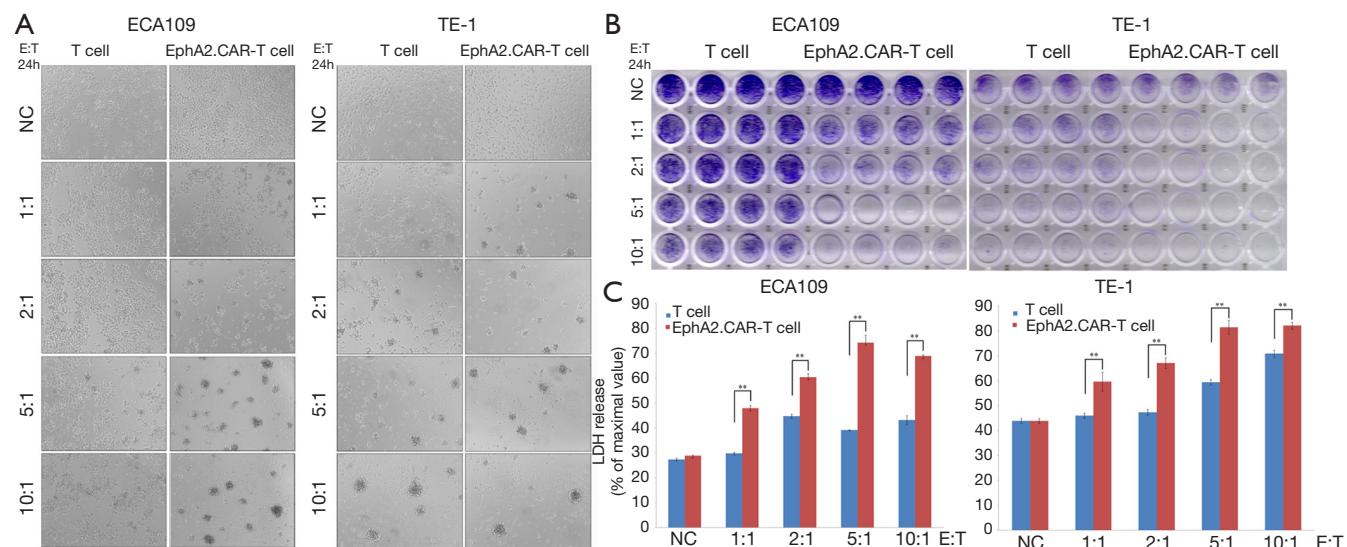


Figure 3 Detection of efficacy of EphA2.CAR-T cells in ESCC cell lines ECA109 and TE-1. The effects of EphA2.CAR-T cells and non-transduced T-cells in killing the ECA109 and TE-1 cells was observed by inverted microscope and measured by Crystal violet staining assay. (A) The image of the ECA109 and TE-1 co-cultured with EphA2.CAR T cells and non-transduced T-cells with different E: T ratio using the inverted microscope; (B) the photograph of crystalline violet staining assay of the ECA109 and TE-1 co-cultured with EphA2.CAR-T cells and non-transduced T-cells with different E: T ratio; (C) detection of the LDH release in the supernatant of the ECA109 and TE-1 co-cultured with EphA2.CAR-T cells and non-transduced T-cells with different E: T ratio after 24 h. *, P<0.05.

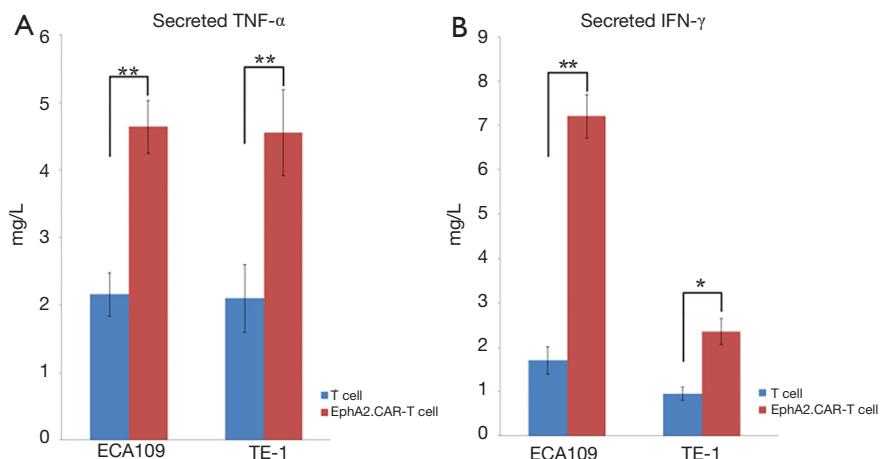


Figure 4 Detection of tumor necrosis factor- α (TNF- α) and interferon-gamma (IFN- γ). TNF- α and IFN- γ secreted in the supernatant of the ECA109 and TE-1 co-cultured with EphA2.CAR-T cells and non-transduced T-cells with different E: T ratio after 24 h was analyzed by ELISA. *, P<0.05; **, P<0.01.

regulating the body's immune status and achieve (27). Cytokines, tumor vaccine, immune checkpoint blockade immunotherapeutic agent (28) and adoptive cell therapy (ACT) has been represented in clinical application and demonstrated great clinical curative effects. The technology of ACT was transferring autologous or allogeneic immune cells to patients after these cells amplification *in vitro*. The transferred immune cells could directly kill tumor cells and mobilize the body immune function.

The current commonly used anti-tumor effector cells could be divided into two categories: the first category was nonspecific effector cell, including autologous lymphokine activated killer cells, cytokine induced killer cells (29) and natural killer cells (30), another type was specific effector cells, including CAR-T cell. The basic characteristics of CAR-T cell was cell killing specifically via recognizing specific TAA (31,32).

EphA2 overexpression induces carcinogenic effects including oncogenesis (33), EMT transformation (34), vasculogenesis (23) and cell growth (35). The unique dual role of EphA2 in tumorigenesis and angiogenesis made it one of the most attractive targets for tumor therapy. And it was reported that EphA2.CAR-T cells were able to recognize and kill EphA2-positive glioma cells *in vitro* and induced GBM tumor regression *in vivo* (20). So, at first, we detected EphA2 expression in ESCC tissues and adjacent tissues via immunofluorescence and immunohistochemistry staining. These results proved that EphA2 expression was up-regulated in ESCC tissues and expressed on the

membrane of ESCC cells. Therefore, EphA2 could be served as TAA of CAR for ESCC treatment.

After the successful construction of EphA.CAR, we firstly considered the tumor cells killing by EphA.CAR-T cells using co-culture with target ECA109 and TE-1 cells *in vitro*. The T cells and EphA2.CAR-T cells were observed as dose-dependent cell killing at E:T ratio of 1:1 to 10:1 for 24 h under an inverted microscope. The efficacy of cell killing of EphA2.CAR-T cells on ECA109 and TE-1 cells was better than that of T cells. Crystalline violet staining assay and LDH release assay was applied to further confirm the cell killing of EphA2.CAR-T cells. The both results were similar to the result observed under an inverted microscope. It was concluded that EphA2.CAR-T cells demonstrated a dose-dependent and specific cell killing *in vitro*.

TNF- α and IFN- γ are cytokines both with anti-tumor characteristics (36,37). The production of TNF- α and IFN- γ by EphA2.CAR-T cells after co-culture with ESCC cells was tested by ELISA. The release levels of TNF- α and IFN- γ in EphA2.CAR-T cells were notably raised. EphA2.CAR-T cells played a vital role in cell killing via increasing cytokines release. The successful cloning of EphA2 scFv into the CAR to the antigen could activate the intracellular signaling transduction of T cells to produce cytokines TNF- α and IFN- γ .

In summary, we firstly successfully constructed the second generation of EphA2.CAR and established EphA2.CAR-T cells. The EphA2.CAR-T cells showed the better efficacy of ESCC cell killing and production of cytokines

in vitro. But, there was a limitation that there was no data of clinical cases or *in vivo* examination in this study. Next, we will test the effect of anti-tumor of EphA2.CAR-T cells *in vivo* and prepare for clinical trial. These findings open a new way for treatment of ESCC by immunotherapy in the future.

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Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

Ethical Statement: The research was authorized by the Ethics Committee of the Affiliated Hospital of Nantong University (No. 2015-070). All patients agreed to use their tissues for scientific research.

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