Association between \textit{RUNX3} promoter methylation and non-small cell lung cancer: a meta-analysis

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\textbf{Background:} Runt-related transcription factor 3 (\textit{RUNX3}) is a known regulator in the transforming growth factor (\textit{TGF})-\textit{β} signaling pathway, which promoter methylation playing a crucial role in diverse neoplasias. However, the relationship between \textit{RUNX3} promoter methylation and non-small cell lung cancer (NSCLC) remains to be clarified.

\textbf{Methods:} We searched Pubmed, Embase, Cochrane Central, and Chinese Biological Medicine database, for articles published in English or Chinese until March 7, 2014. Our main analyses were focused on the association between \textit{RUNX3} promoter methylation and risk of NSCLC by meta-analysis methods. If heterogeneity was observed, we used random effects model to calculate the overall odds ratios, otherwise fixed effects model was used. Subgroup analyses and meta-regression analyses were employed to detect the sources of the heterogeneity. Sensitivity analysis was performed to evaluate the stability of our studies. A funnel plot and Egger's test were conducted to investigate any potential publication bias.

\textbf{Results:} A total of 1,368 samples from 13 literatures were involved in this meta-analysis. The pooled odds ratio (OR) of \textit{RUNX3} methylation in NSCLC specimens compared to non-cancer controls was 6.70 [95% confidence interval (CI): 4.64-9.67]. In the analysis of specimen-types subgroup, the summary OR was 5.79 (95% CI: 3.97-8.46) for tissue specimen subgroup, and that was 45.64 (95% CI: 5.89-353.72) for serum specimen subgroup. The ORs for the age $\leq$ 60 years, 60-65 years and $>$65 years subgroup were 5.19 (95% CI: 3.27-8.24), 9.45 (95% CI: 2.45-36.45) and 13.23 (95% CI: 5.59-31.28) respectively. The result of meta-regression indicated that age was fundamental source of heterogeneity (coefficient =0.61, P=0.046, adjusted $R^2$ =100%). No publication bias was detected. In cancer specimens, the \textit{RUNX3} methylation was associated with histological type of the NSCLC, but no significant differences were found for \textit{RUNX3} methylation in relation to gender, smoking history, tumor TNM stage or tumor differentiation level.

\textbf{Conclusions:} This meta-analysis of pooled data provides additional evidence to support a strong association between methylation of the \textit{RUNX3} promoter and NSCLC. \textit{RUNX3} methylation was increasing with age.

\textbf{Keywords:} Non-small cell lung cancer (NSCLC); runt-related transcription factor 3 (\textit{RUNX3}); promoter; methylation; meta-analysis

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\textbf{Background}:

Lung cancer is one of the most common malignancies and a leading cause of cancer-related deaths worldwide (1). Non-small cell lung cancer (NSCLC) accounts for 80% of all primary lung cancers, which comprises adenocarcinoma, squamous cell carcinoma and large cell carcinoma (2). Current knowledge regarding epigenetic changes play an integral role in the transformation, promotion and progression of cancer (3,4). DNA methylation is one of the most common forms of epigenetic modification. The
abnormal hypermethylation patterns of promoter site in various tumor suppressor genes (TSGs) are a pivotal mechanism in a wide variety of malignancies including lung cancer (5,6).

In humans 1p36.1 chromosomal loci, where RUNX3 is located at this locus, observed to undergo frequent deletion could induce pulmonary carcinogenesis (7,8). RUNX3 is a known regulator in the transforming growth factor (TGF)-β signaling pathway, which has recently been reported as a candidate tumor suppressor (9-11). Decreased RUNX3 expression or deletion are mainly due to methylation or allelic loss, that results in the limited function of Smad proteins and the promotion of TGF-β signaling, which leads to tumor development (12). Previous studies have demonstrated RUNX3 promoter methylation playing a crucial role in neoplasias, including colorectal (13), gastric (14,15), lung (16), bladder (17), breast (18,19) oral (20), and liver cancers (21), either using cell lines, or primary cancer tissues. However, the relationship between RUNX3 promoter methylation and NSCLC remains to be clarified.

Although this association has been investigated in separate studies, the results are somewhat contradictory (22,23), possibly due to small sample size and underpowered in a single study. Therefore, we performed a meta-analysis using all available related studies to assess the association of RUNX3 promoter methylation and NSCLC.

Methods

Search strategies and selection criteria

We searched Pubmed, Embase, Cochrane Central, and Chinese Biological Medicine database, for articles published in English or Chinese. We identified the publications using the text words (RUNX3 or PEBP2αC or CBFA3 or AML2) AND (lung or pulmonic) AND (cancer or neoplasm). The search updated on March 7, 2014. In addition, we also reviewed the reference from retrieved papers and relevant review articles. We only recruited data from fully published papers, not meeting or conference abstracts.

Study selection

Two investigators (Yali Liang and Lianping He) first independently screened the titles and abstracts to identify relevant articles. A second screening was based on full-text articles to further see whether they had met the inclusion criteria. Discrepancies were resolved by consensus.

Studies were included if they meet the following criteria: (I) the specimens from peripheral serum or surgically respected primary tumor tissue (not cell line or sputum); (II) the exposure of interest was RUNX3 promoter methylation; (III) the outcome of interest was NSCLC; (IV) odds ratio (OR) with corresponding 95% confidence intervals (CIs) (or data to calculate them) were published.

Quality assessment

Two investigators independently assessed methodological quality of eligible studies with the Newcastle-Ottawa scale (NOS). The quality scale consists of three parameters: selection, comparability, and exposure assessment. The quality score ranges from 0 to 9. Studies with a score equal to or higher than 4 were considered “high-quality”, whereas those scored less than 4 were considered “low-quality”.

Data extraction

In order to control the bias and improve the reliability, the investigator followed a standardized data-collection form to extract all data. The following information was collected from each study: first author, year of publication, country of the study objects, specimen origin, number of cases and controls, the methylation status of RUNX3 promoter in cancer and control samples, correlation between methylation and clinicopathological characteristics in NSCLC.

Statistical analysis

Our main analyses were focused on the association between RUNX3 promoter methylation and risk of NSCLC. The effect measures of interest were ORs and corresponding 95% CI for case-control study.

Heterogeneity test for pooled ORs was performed by I² statistic (statistically significant level at I² ≥50%) (24). If heterogeneity was observed, we used random effects model (DerSimonian-Laird method) to calculate the overall odds ratios, otherwise fixed effects model (Mantel-Haenszel) was used. Subgroup analysis was performed according to specimen types (lung tissue or peripheral serum), and age categories (≤60 years; between 60 and 65 years; >65 years). If the heterogeneity was strong, meta-regression analyses were employed to analyze the sources of the heterogeneity. Sensitivity analysis was performed by deleting one study in each turn to evaluate the stability.
of the final results. A funnel plot and Egger’s test were conducted to investigate any potential publication bias. We also assessed the correlation between methylation status and clinicopathological characteristics (gender, smoking history, tumor stage, differentiation and histopathology) in NSCLC; histopathological tumor type includes adenocarcinoma, squamous cell carcinoma, and other histological type (large cell carcinoma and mixed histologies carcinoma).

The statistical analyses were performed with Stata12.0 software and review manager 4.2, two-sided P values less than 0.05 were considered statistically significant.

Results

Study selection and characteristics

One hundred and four potentially relevant studies were identified by the electronic search strategy, and 1 study was obtained by manually searching all references cited in the original studies. We justified eligible studies by further screening of their titles, abstracts and full texts. As a result, we retrieved 21 potentially relevant articles. Eight studies were excluded, because one studies did not exactly define as NSCLC (25); one study measured RUNX3 methylation status by the RT-PCR (22); and one study data there had errors (26); and four studies did not establish control groups (27-30). Finally, the remaining 13 studies included in our study (Figure 1) (23,31-42) were included in our meta-analysis. All of them were case-control studies, and two papers were written in Chinese. The main characteristics of the reviewed studies are showed in Table 1. The total sample size was 1,368 (759 cases and 609 controls).

RUNX3 promter methylation and risk of NSCLC

Among these 13 studies, substantial heterogeneity was not obvious ($I^2=47.4\%$). Hence, fixed effects model (Mantel-Haenszel) was used to calculate the pooled OR and 95% CIs (Figure 2). Overall, the pooled OR for RUNX3 methylation in cancer specimens compared with normal specimens was 6.70 (95% CI: 4.64-9.67).

Subgroup analysis and meta-regression

Subgroup analysis was performed according to specimen types (tissue or peripheral serum) and age categories (≤60 years; between 60 and 65 years; >65 years). In the analysis of specimen-types subgroup, the summary OR was 5.79 (95% CI: 3.97-8.46) for tissue specimen subgroup, and that was 45.64 (95% CI: 5.89-353.72) for serum specimen subgroup. There was no evidences of heterogeneity in different specimen types subgroup ($I^2=0\%$). The OR for the age ≤60 years subgroup was 5.19 (95% CI: 3.27-8.24), that for the 60-65 years group was 9.45 (95% CI: 2.45-36.45) and that for the >65 years subgroup was 13.23 (95% CI: 5.59-31.28). Heterogeneity was not observed within different age categories subgroup ($I^2=39.8\%$; $I^2=43.8\%$;
I^2 = 0\%), as showed in Figures 3 and 4.

Heterogeneity was borderline among these 13 studies (I^2 = 47.4\%). Therefore, we performed further analyses to detect the sources of the heterogeneity using the meta-regression method with restricted maximum likelihood modification. The result of meta-regression indicated that the trend in ORs was correlated with age, which accounted for the heterogeneity (coefficient = 0.61, P = 0.046, adjusted R^2 = 100\%). However, the other factor (specimen type) could not explain the heterogeneity. The results are showed in Table 2.

Sensitivity analysis and publication bias
We conducted sensitivity analysis to assess the stability of the overall effects by deleting a single study, the overall ORs did not substantially changed, with a range from 6.00 (95\% CI: 4.13-8.73) to 7.84 (95\% CI: 5.21-11.79).

The funnel plot and Egger's test were performed to estimate publication bias (Figures 5, 6), there was no evidence of publication bias with regard to RUNX3 methylation in relation to NSCLC risk (Egger's test: t = 2.12, P = 0.058).

Correlation between methylation of RUNX3 promoter and clinicopathological characters
Table 3 showed the correlation between methylation of RUNX3 promoter and clinicopathological characters. A portion of these 13 studies provided the relationship of RUNX3 methylation and clinicopathological

| Table 1 Characteristics of studies in this meta-analysis |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Author (Ref)    | Mean/median age (years) [range] | M+/M– | Specimen types | Methods | Methylation site | RUNX3 expression | Quality score |
| Hou DR et al. (34) 2009, China | 55 [38-64] | 30/32 | 11/51 | Tissue | MSP | Promoter hypermethylation | Negative | 6 |
| Zhang Y et al. (31) 2011, China | 59 [35-80] | 18/60 | 8/70 | Tissue | MSP | Cpg islands | Not reported | 5 |
| Yu GP et al. (32) 2012, China | 57 [38-72] | 26/32 | 10/48 | Tissue | MSP | Promoter | Not reported | 7 |
| Lu DG et al. (33) 2011, China | 59.6 [42-75] | 25/37 | 0/46 | Serum* | MSP | Promoter hypermethylation | Not reported | 5 |
| Yanagawa N et al. (35) 2003, Japan | 67.3 [39-86] | 15/60 | 2/73 | Tissue | MSP | Promoter hypermethylation | Not reported | 6 |
| Suzuki M et al. (36) 2005, Japan | 65 | 25/92 | 0/51 | Tissue | MSP | Cpg islands | Not reported | 4 |
| Yanagawa N et al. (37) 2007, Japan | 68.1 [39-86] | 25/76 | 3/98 | Tissue | MSP | Promoter hypermethylation | Not reported | 8 |
| Yoshino M et al. (38) 2009, Japan | 63.2 [44-90] | 9/35 | 2/30 | Tissue | MSP | Cpg islands | Not reported | 8 |
| Li QL et al. (39) 2004, Korea | Not reported | 6/19 | 0/25 | Tissue | MSP | Promoter | Not reported | 7 |
| Chung JH et al. (40) 2011, Korea | 59.2 [34-85] | 29/61 | 0/20 | Tissue | q-MSP | Cpg islands | Not reported | 8 |
| Tan SH et al. (23) 2007, Singapore | Not reported | 11/9 | 0/10 | Serum* | MSP | Promoter hypermethylation | Not reported | 4 |
| Omar MF et al. (41) 2012, Singapore | Not reported | 3/6 | 3/2 | Tissue | MSP | Promoter hypermethylation | Positive | 4 |
| Licchesi JD et al. (42) 2008, USA | 69.6 [48-80] | 17/1 | 13/33 | Tissue | MSP | Promoter hypermethylation | Negative | 6 |

M+, the number of tissues with methylation; M–, the number of tissues with unmethylation. *, peripheral serum.
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**Figure 2** Forest plot of runt-related transcription factor 3 (RUNX3) promoter methylation in cancer specimens vs. non-cancer controls.

**Figure 3** Forest plot of different specimen types subgroups analysis.

characteristics in their cancer specimens. The overall results demonstrated that RUNX3 methylation was less frequent in adenocarcinoma compared with squamous cell carcinoma (OR = 2.25, 95% CI: 1.48-3.42), but the frequencies was similar in adenocarcinoma and other histological type (OR = 0.49, 95% CI: 0.22-1.10). Highly and moderately differentiated cancer specimens also had a lower methylation than poor differentiation (OR = 0.39, 95% CI: 0.06-2.36).

There were no significant differences in RUNX3
Figure 4 Forest plot of different age categories subgroups analysis.

Table 2 The results of meta-regression

<table>
<thead>
<tr>
<th>Sources</th>
<th>Coefficient (95% CI)</th>
<th>t</th>
<th>P</th>
<th>Adjusted R² (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Specimen type</td>
<td>-2.82 (−6.29-0.64)</td>
<td>−1.93</td>
<td>0.096</td>
<td>58.78</td>
</tr>
<tr>
<td>Age</td>
<td>0.61 (0.01-1.20)</td>
<td>2.42</td>
<td>0.046</td>
<td>100.00</td>
</tr>
</tbody>
</table>

CI, confidence interval.
methylation status in cancer sample in relation to gender, smoking and tumor stage. The results are shown in Figures 7-12.

**Discussion**

Our meta-analysis focuses on relationship between \textit{RUNX3} promoter methylation and the risk of NSCLC. The overall OR for methylation status in cancer versus normal specimens was 6.70 (95% CI: 4.64-9.67) by a fixed effects model on pooled data from 13 studies. In the specimen types-specific subgroup analysis, results showed: the OR in the tissue sample subgroup was 5.79 (95% CI: 3.97-8.46), that in serum samples subgroup was 45.64 (95% CI: 5.89-353.72), which further confirmed \textit{RUNX3} methylation was a potential risk factor for NSCLC. Among peripheral blood of non-cancer objects, the methylation of \textit{RUNX3} was absent, and so the methylation of \textit{RUNX3} could be regarded as cancer-specific phenomenon. Therefore, in terms of clinical application, the detection \textit{RUNX3} methylation of peripheral blood may be useful as a diagnostic approach. The trend of association between \textit{RUNX3} methylation and NSCLC was correlated with age. The OR was 5.19 (95% CI: 3.27-8.24) for the age ≤60 years subgroup, 9.45 (95% CI: 2.45-36.45) for the group of 60-65 years, and 13.23 (95% CI: 5.59-31.28) for the >65 years subgroup. The coefficient for age was calculated to be 0.61 by meta-regression analysis, indicating that the tendency for \textit{RUNX3} methylation increased with age. DNA methylation, genomic imprinting, and histone modifications were examples of epigenetic factors known to undergo change in the aging and malignant counterparts (43). \textit{RUNX3} exhibited altered DNA methylation patterns in aging, displaying sometimes tissue- and cell type-specific features with consequent different functional outcomes (44). Some studies reported \textit{RUNX3} methylation occurring preferentially in older malignant tumor patients (45). These results suggested that \textit{RUNX3} methylation related with individual age in malignancies. We found that the ORs for \textit{RUNX3} methylation increased from 5.19 in the younger age group, through 9.45, to 13.33 in the oldest age group. The coefficient for age was calculated to be 0.61 by meta-regression analysis, indicating that the tendency for \textit{RUNX3} methylation increased with advancing age. The results suggested \textit{RUNX3} methylation may be preferentially occurs in older NSCLC patients.

Although no publication bias was detected using Egger’s test, the funnel plot showed one of the studies exceeded the 95% confidence limits. We performed sensitivity analysis in each to estimate the robustness of our results by deleting one study. The overall ORs were slightly changed from 6.00

<table>
<thead>
<tr>
<th>NO. study</th>
<th>Characters</th>
<th>M+/M–</th>
<th>OR (95% CI)</th>
<th>I² (%)</th>
<th>P (Egger’ test)</th>
</tr>
</thead>
<tbody>
<tr>
<td>6</td>
<td>Gender</td>
<td>Male</td>
<td>73/156</td>
<td>0.80 (0.51-1.26)</td>
<td>8.0</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Female</td>
<td>57/116</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>Smoking</td>
<td>Yes</td>
<td>52/133</td>
<td>0.64 (0.38-1.07)</td>
<td>25.2</td>
</tr>
<tr>
<td></td>
<td></td>
<td>No</td>
<td>53/102</td>
<td></td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>Pathological type</td>
<td>ACC/SCC</td>
<td>102/201; 44/180</td>
<td>2.25 (1.48-3.42)</td>
<td>34.3</td>
</tr>
<tr>
<td>5</td>
<td>ACC/other type</td>
<td>46/124; 18/18</td>
<td>0.49 (0.22-1.10)</td>
<td>0</td>
<td>0.595</td>
</tr>
<tr>
<td>5</td>
<td>SCC/other type</td>
<td>21/106; 18/9</td>
<td>0.10 (0.04-0.29)</td>
<td>0</td>
<td>0.068</td>
</tr>
<tr>
<td>3</td>
<td>Differentiation</td>
<td>H/M</td>
<td>40/68</td>
<td>0.39 (0.06-2.36)</td>
<td>86.4</td>
</tr>
<tr>
<td></td>
<td></td>
<td>P</td>
<td>41/33</td>
<td>0.47 (0.26-0.85)</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>TNM stage</td>
<td>I-II</td>
<td>58/151</td>
<td>0.79 (0.24-2.62)</td>
<td>78.3</td>
</tr>
<tr>
<td></td>
<td></td>
<td>III-IV</td>
<td>69/105</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

M+, the number of tissues with methylation; M–, the number of tissues with unmethylation; other type, large-cell carcinoma and adenosquamous cell carcinoma. H/M, highly and moderately differentiated; P, poor differentiation. \(^\text{a}\) by random effect model (DerSimonian-Laird method); \(^\text{b}\) by fixed effects model (Mantel-Haenszel). Abbreviations: RUNX3, runt-related transcription factor 3; SCC, squamous cell carcinoma; ACC, adenocarcinoma; NSCLC, non-small cell lung cancer; OR, odds ratio; CI, confidence interval.
Figure 7 Forest plot of smoking in non-small cell lung cancer (NSCLC).

Figure 8 Forest plot of tumor stage in non-small cell lung cancer (NSCLC).

Figure 9 Forest plot of differentiation in non-small cell lung cancer (NSCLC).
Figure 10 Forest plot of different histological type (ACC/SCC) in non-small cell lung cancer (NSCLC). ACC, adenocarcinoma; SCC, squamous cell carcinoma.

Figure 11 Forest plot of different histological type (ACC/Other) in non-small cell lung cancer (NSCLC). ACC, adenocarcinoma.

Figure 12 Forest plot of different histological type (SCC/Other) in non-small cell lung cancer (NSCLC). SCC, squamous cell carcinoma.
(95% CI: 4.13-8.73) to 7.84 (95% CI: 5.21-11.79), which were still significant. To produce a more robust estimation, we performed sensitivity analysis by deleting one study in each turn. The overall ORs were slightly changed from 6.00 (95% CI: 4.13-8.73) to 7.84 (95% CI: 5.21-11.79), which were still significant, indicating a strong association between RUNX3 promoter methylation and NSCLC.

There were no significant differences in RUNX3 methylation in cancer tissues in relation to gender, smoking history, or tumor TNM stage.

The aggregated results found that RUNX3 methylation was less frequent in squamous cell carcinoma compared with adenocarcinoma and other histological type, suggesting that inactivation of RUNX3 might play a less significant role in the pathogenesis of squamous cell carcinoma, as those previous studies suggested (30).

Analysis of the pooled data also showed that undifferentiated NSCLC had a higher frequency of promoter methylation than well-differentiated, which was significant in the fixed model (OR =0.47, CI: 0.26-0.85) but non-significant in the random effects model. This phenomenon may be associated with the smaller number of studies analyzed. But the results also indicated that RUNX3 promoter methylation may be related to poor prognosis.

Some limitations of this meta-analysis should be addressed. First, Although we searched literature as completely as possible, the results calculated in our meta-analysis maybe exist bias as we only collected full published papers and articles published in English or Chinese. Second, our results were based on unadjusted, whereas a more precise analysis should be conducted if adjustment estimates were available.

Conclusions

Despite these limitations, this meta-analysis provides additional evidence to support a strong association between methylation of the RUNX3 promoter and NSCLC. The tendency of association for RUNX3 methylation increased with age. The RUNX3 methylation was also associated with histological type of the NSCLC. However, it is necessary to conduct large sample studies using standardized and well-matched controls.

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