Progressive CT findings and positive RT-PCR again of recovered and discharged patients with COVID-19

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As of April 27, 2020, 2,878,196 (85,530 new) COVID-19 patients were confirmed across the world (1), within which 77,555 (82,836 confirmed) recovered in China. Previous studies focused on clinical and radiological features of patients with confirmed infection (2,3), or recovered patients with no symptoms and no changes of follow-up chest computed tomography (CT) findings (4). To our knowledge, both the progressive CT findings and fever again of recovered and discharged patients has not been reported.

One representative hospitalized patient with COVID-19 treated at Tongji Hospital, Wuhan, China, from January 20, 2020, to February 8, 2020 was discharged and isolated at home, because he met the discharge criteria in China: (I) normal temperature for more than 3 days; (II) resolved respiratory symptoms; (III) substantially improved acute exudative lesions on CT images; and (IV) 2 consecutively negative reverse transcription polymerase chain reaction (RT-PCR) test results separated by at least 24 hours (5). However, on February 13, 2020, the patient was admitted to the isolation ward and received supplemental oxygen through nasal cannula once again, owing to symptoms of fever and shortness of breath, and progressive CT findings. RT-PCR test was performed via nasopharyngeal swabs, and severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) nucleic acid was positive on February 14, 2020. The RT-PCR test performed in Tongji Hospital was reasonably reliable, due to the RT-PCR detection reagent used in the clinical laboratory proved by China Food and Drug Administration (CFDA) with 100% positive and negative reference coincidence rate.

Laboratory studies, clinical data, and radiological features were collected from electronic medical records. The local ethical review board approved this study and waived the requirement to obtain individual informed consent.

A 40-year-old patient (male) living in Wuhan, was admitted to a fever clinic on January 20, 2020, with symptoms of fever for 2 days. Chest CT revealed multiple ground-glass opacities (GGOs) distributed in the peripheral and peribronchovascular area of bilateral lungs (Figure 1A). The major axis of maximal opacity was 2.8 cm in right lower lobe. Respiratory tract virus antigen and antibody tests, including cytomegalovirus, influenza virus A and B, human parainfluenza virus, respiratory syncytial virus and adenovirus, showed all negative results on January 22, 2020, and SARS-CoV-2 nucleic acid was positive one day later. A 4-day follow-up CT showed multiple GGOs enlarged significantly with multifocal consolidation, interlobular septal thickening, vessel thickening and air bronchogram, which involved all pulmonary segments (Figure 1B). The major axis of maximal lesion was 7.5 cm in right lower lobe. On January 27, 2020, the 2nd follow-up CT illustrated multiple GGOs further enlarged (the major axis of maximal lesion was 10.1 cm in right lower lobe), but consolidation
partial absorbed (Figure 1C). After 7 days, the 3rd follow-up CT represented remarkable absorption of multiple GGOs in bilateral lungs and multiple linear consolidation (Figure 1D). The patient remained negative after 2 consecutive RT-PCR tests on February 4 and 6, 2020. The time from symptom onset to recovery was 22 days.

After hospital discharge, the patient was isolated at home for 5 days. On day 27 of illness, the patient had a fever again (38.2 °C), and CT showed some GGOs absorbed but others progressed to multifocal organizing consolidation in bilateral lungs (Figure 1E). Positive RT-PCR test results and laboratory studies suggested inflammatory situation, with lymphocyte count 0.70 (normal: 1.10–3.20)×10⁹/L, erythrocyte sedimentation rate 68 (normal: 0–15) mm/H, procalcitonin 0.13 (normal: 0.02–0.05) ng/mL, interleukin-2 receptor 1,108 (normal: 223–710) U/ml, interleukin-6 9.97 (normal: <7) pg/mL, interleukin-10 10.2 (normal: <9.1) pg/mL, and tumor necrosis factor 15.7 (normal: <8.1) pg/mL. From February 13, 2020, the patient was admitted to the isolation ward and received antibiotic, antiviral, hormone, and stomach-protecting therapies once again.

On February 18, 2020, the follow-up CT showed some consolidation enlarged in left lower lobe and right upper lobe, while other linear consolidation remained (Figure 1F). Moreover, laboratory studies matched the progressive CT findings, with ferritin 1,846.5 (normal: 30–400) μg/L, procalcitonin 0.13 (normal: 0.02–0.05) ng/mL, interleukin-2 receptor 1,108 (normal: 223–710) U/ml, interleukin-6 9.97 (normal: <7) pg/mL, interleukin-10 10.2 (normal: <9.1) pg/mL, and tumor necrosis factor 15.7 (normal: <8.1) pg/mL. From February 13, 2020, the patient was admitted to the isolation ward and received antibiotic, antiviral, hormone, and stomach-protecting therapies once again.

On February 29, 2020, the LDH (162 U/L), procalcitonin (0.03 ng/mL), and lymphocyte count (1.28×10⁹/L) recovered, RT-PCR tests were negative, and CT scan showed multiple consolidative opacities were partially absorbed in bilateral lungs. Two days later, the serological SARS-CoV-2 antibody test was positive with IgM 19.27 (normal: ≤10) AU/mL and SARS-CoV-2 IgG 24.68 (normal: ≤10) AU/mL, and ferritin 630.6 μg/L elevated significantly. The patient was still in the hospital for treatment because of high antibodies and ferritin.

A patient with COVID-19 who satisfied the criteria for discharge (absence of clinical symptoms, substantial recovery) was retested positive for COVID-19. This demonstrates the necessity for thorough follow-up and monitoring of discharged patients.
improvement of acute exudative lesion on CT images, and 2 consecutively negative RT-PCR test results) (5) had a fever, processive CT findings and positive RT-PCR test results 5 days later. These findings indicated that at least a proportion of recovered patients might still carry virus, though his SARS-CoV-2 nucleic acid tests were negative through twice nasopharyngeal swabs. We speculated that SARS-CoV-2 was still active in lower respiratory tract, which was also supported by multiple GGOs and linear consolidation on CT images.

Noticeably, 3 days before discharge, the procalcitonin (0.04 ng/mL) and lymphocyte count (1.27×10^9/L) recovered but LDH (313 U/L) was still significantly high. This suggested that the pneumonia had not been completely absorbed. On February 28, the press conference of the joint prevention and control mechanism of the state council in China, reported a few discharged cases with positive RT-PCR tests from different provinces. It indicated that current criteria for hospital discharge could be reevaluated.

Our finding suggested that current discharge criteria could take into account some laboratory studies, such as interleukin6, LDH, and SARS-CoV-2 IgM and IgG. In the future, we will investigate longitudinal studies on a large cohort, which can help understand chest morphometry of COVID-19 and provide a useful path towards improvement of clinical strategies against the disease.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/jtd-20-1417). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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