Pre-analytic variability in cardiovascular biomarker testing

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Abstract: The impact of laboratory medicine on clinical cardiology has dramatically increased over the years and a lot of cardiovascular biomarkers have been recently proposed. In order to avoid clinical mistakes, physicians should be well aware of all the aspects, which could affect the quality of laboratory results, remembering that pre-analytic variability is an often overlooked significant source of bias, determining the vast majority of laboratory errors. This review addresses the determinants of pre-analytical variability in cardiovascular biomarker testing, focusing on the most widespread biomarkers, which are cardiac troponins and natriuretic peptides.

Keywords: Laboratory variability; biomarker; cardiology

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Introduction

The impact of laboratory medicine on clinical cardiology has dramatically increased over the years (1,2) and a lot of cardiovascular biomarkers have been recently proposed. However, the most widespread biomarkers in cardiology are cardiac troponins and natriuretic peptides, on which this review will focus.

Cardiac troponin I (cTnI) and T (cTnT) have become the most reliable tool for diagnosing acute myocardial infarction (3). According to recent evidence, serial testing with assessment of troponins’ kinetic are recommended if acute cardiac problems are suspected. With the introduction of high sensitivity cardiac troponin assays, quantification of cTn levels is nowadays possible in almost everyone and cTn’s elevation is linked to adverse events (4). cTn rises mainly in acute coronary syndromes but also in chronic or acute renal impairment, hearth failure, sepsis, hypertensive crisis, arrhythmias, pulmonary embolism, myocarditis, stroke and other cardiac problems (5,6).

Nowadays laboratory test could also investigate the function of the heart, especially its endocrine activity, through the family of peptide hormones with potent natriuretic activity such as type B natriuretic peptide (BNP), atrial natriuretic peptide (ANP), type C natriuretic peptide, urodilatin and dendroaspis natriuretic peptide (7).

Usually clinical laboratories concentrate on BNP and N-terminal (NT)-proBNP, which derive from proBNP, the common precursor peptide (1). The European Society of Cardiology Task Force for the diagnosis and treatment of chronic heart failure suggests a close relation between natriuretic peptides levels and heart failure. In fact there is a progressive increase of BNP levels from patients with normal heart function, to patients with preclinical alterations (8) or overt heart failure. Normal levels in untreated patients have a strong negative predictive value for the presence of heart failure and Guidelines recommend testing BNP or NT-proBNP levels in symptomatic patients in order to rule out the diagnosis of heart failure (9).
Laboratory diagnostics

Laboratory diagnostics is commonly defined as the act or process of identifying the nature and cause of a disease by means of *in vitro* diagnosis testing. Laboratory diagnostics is traditionally divided into three different phases: pre-analytical, analytical and post-analytical. The pre-analytical phase represents the different activities necessary to obtain the biological specimens on which to perform the laboratory evaluations and most of these activities are located outside the clinical laboratory (patients preparation, sample collection, handling and transportation, preparation and storage of the specimens). The leading causes of pre-analytical variability are related to patient preparation, blood drawing, sample transportation and preparation, as reported in Table 1 (10). Several lines of evidence attest that the vast majority of laboratory errors (i.e., approximately 70%), derives from pre-analytical rather than analytical or post-analytical phases (11).

The receipt of unsuitable sample is relative common in laboratory practice and represents a problem for tests’ quality and results and, consequently, for patients safety. Detection and management of unsuitable samples are necessary to avoid dangerous clinical consequences as recently underlined (12).

Along samples unsuitable for quality (haemolyzed or clotted) and quantity (insufficient or inappropriate volume) (13), additional mishandling practices such as prolonged venous stasis (14), inappropriate mixing of primary blood tubes (15), inappropriate condition of transportation and storage of specimens (16), different conditions of centrifugations like the time (17) and the use or not of centrifuge brake (18) represent a possible cause of inaccuracy of results.

The impact of the blood collection devices (parts of vial, gel separator or tube stopper) on clinical chemistry assay is an intriguing and less known aspect, which can alter laboratory results, as recently reported (19,20).

Even the possible interference of medical contrast media on laboratory testing should be taken into account as an important source of analytical inaccuracy (21,22). All the above mentioned aspects may influence the huge number of tests performed in clinical laboratory.

Patient’s conditions

Patient conditions (i.e., fasting status, exercise, posture and circadian changes) are often an overlooked source of bias in laboratory diagnostics (10).

Fasting status

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Lipemia is the second (after hemolysis) most frequent cause of interference in various laboratory methods. Inadequate time of blood sampling after the meal is the most common pre-analytical cause of lipemia and therefore current recommendations entails avoiding foods and beverages for 6-12 hours before testing (23), even if lipemia can be removed in most cases with different techniques like ultracentrifugation or the use of solvents such as LipoClear®. A light meal does not influence the laboratory coagulation tests while significant variation of several clinical chemistry and hematological parameters was described (24-26). At this regard no data are available for cTn, where LipoClear® is not suitable for lipemia removal (27) having shown unacceptable recovery for TnT, according to the desirable specification for imprecision criteria. High speed centrifugation should be used for lipemia removal when proceeding to cardiac troponins’ determination in lipemic specimens (28).

Physical activity

Physical activity is an important pre-analytical variable.

### Table 1 The leading causes of pre-analytical variability

<table>
<thead>
<tr>
<th>Patient preparation</th>
<th>Fasting status</th>
<th>Exercise</th>
<th>Posture</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood drawing</td>
<td>Misidentification</td>
<td>Insufficient volume</td>
<td>Hemolysis</td>
</tr>
<tr>
<td>Contamination</td>
<td>Venous stasis</td>
<td>Blood collection devices</td>
<td>Mixing</td>
</tr>
<tr>
<td>Sample transportation and preparation</td>
<td>Time, temperature, integrity</td>
<td>Centrifugation</td>
<td></td>
</tr>
</tbody>
</table>

Middle and long-term endurance and/or strenuous exercise trigger transient elevations of cardiac biomarkers such as cTn, natriuretic peptides and several others biomarkers (29-31). Increased levels of cTnI and cTnT were measured respectively in 30-90% and 10-100% endurance athletes after exercise, being the cut-off for normality more often exceeded when late generation assays were used (30,32,33). Also normal physical activity can induce transient cTn elevation in almost 50% of healthy subjects. These results were obtained with previous generation immunoassay cTnT and cTnI in adolescents after basketball training (34) or treadmill running (35) and confirmed with high-sensitivity immunoassay cTnT after maximal bicycle stress test (36) or recreational resistance training like kettlebell workout (37).

NT-proBNP levels are also affected by vigorous physical activity, resulting much higher in ultramarathon than in half-marathon runners (38,39). No half-marathon runner exhibited NT-proBNP concentrations exceeding the diagnostic thresholds of this assay, whereas values above the cut-off were observed in nearly one-third of ultramarathon runners (38). The substantial raise of NT-proBNP is a clear consequence of the increased myocardiocyte stretch deriving from strenuous aerobic exercise and is barely influenced by plasma volume change or fluid imbalance (38).

In conclusion several lines of evidence show that physical activity is an important source of pre-analytic variability in cTnT and cTnI testing, especially when latest generation immunoassay are used. Data about the influence of physical activity on BNP determination are lacking but, being BNP cleaved from the same precursor of NT-proBNP, it seems reasonable that also this biomarker may be influenced by exercise.

A great caution in the evaluation of cardiac biomarker determination shortly after strenuous physical activity is therefore advisable.

**Position**

Body position during blood collection may induce significant variations of plasma volume, thereby affecting the results of many laboratory tests. cTnT seems to be independent from posture position (supine, seated, standing position) (40), while data on BNP are lacking.

**Circadian changes**

The cyclic variation showed by many biomarkers (diurnal, monthly, seasonal) is another well-known source of pre-analytical variability. cTnT, measured with a high-sensitivity assay, exhibits a diurnal rhythm, characterized by peak concentrations during the morning hours, gradually decreasing throughout daytime and rising again during nighttime (41), while no data are available for natriuretic peptides.

**Age and gender**

Using high-sensitivity immunoassay, age and gender-dependent effects have been observed both for cTnI (42,43) and cTnT (44,45). This aspect plays an important role in epidemiological studies and in defining the appropriate cardiac 99th percentile of a reference population for clinical purposes, while it is less affecting natriuretic peptides.

**Collection devices**

Observations about the role played by collection tubes in the determination of cTnI and T are discordant. Some authors reported lower cTn levels, when measured with no high sensitivity immunoassay in heparin plasma compared with serum, suggesting that the binding of heparin to troponins could decrease their immunoreactivity, depending on the kind of reaction antibody used (46). Another study did not confirm these observations for cTnI, which was shown not to be influenced by the type of collection tubes used (without anticoagulant, with heparin and gel separator and with heparin but without gel separator) (20). Serum and sodium-citrate plasma appeared also to be interchangeable for cTnI measurement (47).

BNP measurement seemed to be highly dependent on type of collection tube, being levels measured on K2-ethylene diamine tetraacetic acid (EDTA) significantly different if compared to lithium heparin with gel separator. Some studies reported an underestimation in BNP measurements performed in K2-EDTA if compared to lithium heparin (48), but other reported an underestimation in measurements performed in heparin and serum samples compared to EDTA (49). Finally other authors suggested heparin plasma as an attractive alternative to the established EDTA samples for the BNP determination (50) or 60%, 39%, 70% and 48% lower results in collection tubes containing citrate, heparin, fluoride and no anticoagulants respectively, compared with samples collected into EDTA tubes (51). The different results obtained in these studies may be due to the different immunoassay used, even if a role of the biological
matrix has been also suggested (49). For NT-proBNP determinations EDTA or heparinized plasma samples have been proposed to be indifferently used (52).

Standardization of collection tubes is therefore requested in order to avoid a possible source of results variability.

**Haemolysis**

Haemolysis is defined as the presence of free haemoglobin concentrations above 0.3 g/L (18.8 mmol/L), which confers a detectable pink to red color to plasma or serum, visible after centrifugation of the specimen (53).

In *vitro* haemolysis remains the leading cause of unsuitable routine and stat specimens for both outpatient and inpatient samples (54,55). Haemolytic specimens are relative frequent, with a prevalence of 3.3% of all of the routine samples, accounting for 40-70% of all unsuitable specimens.

Haemolysis occurs as a consequence of the release of hemoglobin and other intracellular components from blood cells and may occur both *in vivo* and *in vitro*. In *vivo* haemolysis, due to several pathological conditions, is uncommon and accounts for less than 2% of all haemolytic specimens (56). In *vitro* haemolysis may be due to problems and difficulty during blood collection, or inappropriate handling, storage or centrifugation of the sample (57).

Interference of haemolysis on laboratory testing might be caused by leakage of haemoglobin and other intracellular components into the surrounding fluid, which could induce false elevations of some analyte, dilution effects, chemical interference of free hemoglobin in a variety of analytic reactions and analyte concentration-dependent spectrophotometric interference (57).

Haemolysis interference is less frequently reported in immunoassay than in photometric assay (58). Interference may occur if the reagent antibodies used in immunoassay are poorly specific and cross-react with some of the compounds released from the erythrocyte. Furthermore, some released compound may bind to the analyte and inhibit the reaction with the antibody (59).

Influence of hemolysis on routine clinical chemistry and coagulation testing has been comprehensively evaluated (60,61). However, reliable information on the potential bias on cardiac marker testing arising from *in vitro* haemolysis is lacking or, in some case, controversial (62-64).

In our previous study a moderate blood cells lysis, producing a concentration of free haemoglobin up to 0.6 g/L, was not associated to significant clinical difference in the determination accuracy of routine cardiac biomarkers with the analyzer Modular System E (NT-proBNP, cTnT) and Access 2 (cTnI, BNP, Mioglobulin). Little or moderate sign of haemolysis in the specimens therefore appears not to significantly affect the determination of cardiac biomarkers, if not high sensitivity cTn assays are used (65). On the contrary, higher levels of haemolysis, biased negatively cTnT testing (66). High sensitivity cTnT showed a negative interference with increasing degrees of hemolysis (67,68).

For the current cTnI assay, different degree of interference were shown between different immunoassays, ranging from a positive interference of <1% up to 576% (67).

In conclusion haemolytic specimens may be a variable source of bias in cardiac cTn results. Further studies are needed to evaluate the impact of hemolysis on the very low concentration of the more and more high-sensitivity cTn immunoassay.

**Stability**

Many studies have analyzed BNP and NT-proBNP stability, but their conclusion are discordant (52,69,70) and, once again, this could depend on the different assays used. Without adding protease inhibitors, BNP stability determined with a fully automated micro-particle enzyme immunoassay using frozen plasma is not reliable (70). Using a chemiluminescence immunoassay, different stability of BNP in relation to its plasma concentration was shown: in frozen plasma BNP remained stable only at physiological levels (71,72), while a pathological levels its concentration decreased over time (71). At room temperature EDTA-plasma BNP declined significantly after 4 h and decreased 2-fold after 48 h of storage (70). Similar significant decrease was shown when BNP was stored at 4 °C (70).

NT-proBNP may be stored at 20 °C for at least 4 months in the absence of protease inhibitors, without a relevant loss of the immunoreactive analyte (70) and, in EDTA-plasma, for up to 48 h at room temperature. Same results were obtained storing NT-proBNP for 6 days at 4 °C or 10 days at 20 °C (52).

**Conclusions**

Pre-analytic variability is an important aspect that affects the quality of the laboratory results, included cardiac biomarkers determination. In order to correctly evaluate cardiac biomarkers results physicians, especially cardiologists and laboratory specialists, should be aware of
all the aspects, which could affect the quality of laboratory results, remembering that pre-analytic variability is an often overlooked significant source of bias.

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Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

References

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