Introduction

Porcine bioprosthetic valves have been widely used for mitral valve replacement since the 1960s (1). Several manufacturers have adopted unique strategies and technologies for the design and manufacture of valves to improve hemodynamic performance and durability (2,3). However, porcine valve deterioration depends on the implanted position. The mitral porcine valve is...
located between the left atrium and the left ventricle and is therefore exposed to left ventricular blood flow and pressure. Clinical follow-up studies of patients with bioprosthetic valves have shown that porcine mitral valves still have several problems in terms of freedom from reoperation, survival outcomes, and valve-related morbidity and mortality (4-6). To determine the underlying causes of porcine mitral valve deterioration, the present retrospective study reviewed the morphological and pathological features of porcine mitral valves after second valve replacements at one center.

We present the following article in accordance with the STROBE reporting checklist (available at http://dx.doi.org/10.21037/jtd-20-3578).

Methods

From September 2018 to December 2019, a total of 12 porcine mitral valves were explanted for analysis at our center (Beijing Anzhen Hospital, Capital Medical University, Beijing, China). All patients in this study were provided informed consent for undergoing the procedures. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study design was approved by the Ethics Review Committee of Beijing Anzhen Hospital (No. 2020069x). The baseline characteristics of the patients (gender, age, age at first implantation, implant duration, age at revision surgery, and echocardiography results) and valve explants (valve brand, implantation site, and valve type) were obtained from the medical records. Echocardiography was performed to determine the porcine valve function preoperatively. The causes of valve dysfunction include structural valve deterioration (cusp tears, commissural dehiscence, and calcification), nonstructural valve deterioration (valvular endocarditis and pannus formation), thrombosis, and infection (7,8).

Morphological examination

The explanted deteriorated valves were cleaned with 0.9% stroke-physiological saline solution and preserved with 4% paraformaldehyde. Cusp tears, commissural dehiscence and pannus were identified and classified by morphological examination. Based on the location and manner of leaflet tears, cusp tears were categorized into four types: type I (longitudinal tears involving free edges of the cusp), type II (parallel to the sewing ring), type III (large, round, or oval perforations at the center of the cusp), and type IV (several pinholes in the center of the cusp) (Figure 1) (9). Commisural dehiscence was regarded as separated leaflets from prosthetic stents without cusp tears (10). Pannus was divided into the following types: type I (located on the surface of the cusp), type II (covering <2 mm of the surface of the cusp), type III (covering >2 mm of the surface of the cusp), and type IV (very severe, surrounding the entire surface of the cusp and causing cusp contracture) (9).

Microcomputed tomography scanning

Microcomputed tomography (micro-CT) imaging has been used to identify small calcified tissues of porcine valves with high accuracy (11). All metals, plastics, and fabrics were removed from the explanted valves and only leaflet tissues were examined with micro-CT (Skyscan 1076, Bruker Corporation, MA, USA) to obtain two-dimensional layer-by-layer images. Finally, three-dimensional reconstruction software (MIMICS 14.0, Materialise, Leuven, Belgium) was used to reconstruct the three-dimensional structure of the valve leaflets (12). The calcified parts were marked in red to distinguish between calcified and noncalcified leaflets. It is worth mentioning that all micro-CT scans were performed by experienced personnel blinded to the clinical outcomes. Calcifications were classified as type 0 (no calcification), type I (mild lesions and macrocalcification), type II (moderate nodules <1 mm in diameter), type III (large solitary nodules 1–2 mm in size), and type IV (confluent nodules >2 mm in diameter and capable of puncturing the cusp surface) (9,13).

Pathological examination

Pathological examination was performed using specimen prepared from representative samples of leaflet tissue. Decalcification was necessary prior to pathological examination. All sections were embedded in paraffin, sectioned at 4 μm, and stained with hematoxylin and eosin (H&E) (7), and the Von Kossa’s method.

Statistical analysis

Comparisons of continuous variables are expressed as mean ± standard deviation. The implant duration and patient age at first implantation were analyzed using t-tests. All statistical analyses were performed using SPSS version 26 (IBM Corporation, Armonk, NY, USA). Statistical
significance was set at P<0.05.

Results

A total of 5 Hancock II (Medtronic Heart Valve Division, USA) porcine valves and 7 Carpentier-Edwards Supraannular (CE-SAV; Edwards Lifesciences Corporation, Irvine, CA, USA) porcine valves were explanted from 3 males (25%) and 9 females (75%). The baseline characteristics of the patients are listed in Table 1. The mean age was 55±19.64 years (range, 18–74 years). The mean age at first implantation was 45.42±19.58 years, (range, 11–64 years). The indications for first surgery were rheumatic heart disease in 8 patients (66.67%), infective endocarditis in 2 patients (16.67%), degenerative valvular disease in one patient (8.33%), and congenital heart disease in one patient (8.33%). Four (33.33%) patients took warfarin before the explantation due to atrial fibrillation. The mean implant duration was 9.39±4.14 years (range, 4.25–18.75 years). Structural valve deterioration was found in 11 (91.67%) valves, and nonstructural valve deterioration was found in one patient (8.33%) (Table 2). There were no cases of valve deterioration due to thrombosis or infection.

Overall, 6 patients (50%) showed one or more cusp tears (Figure 2). Only type I tears were found in the valve with cusp tears (Table 2). Commissural dehiscence accounted for valve dysfunction in 5 patients (41.67%) (Figure 2, Table 2).

Table 1 Patient basic characteristics

<table>
<thead>
<tr>
<th>Demographics and Characteristics</th>
<th>Porcine valves</th>
</tr>
</thead>
<tbody>
<tr>
<td>N</td>
<td>12</td>
</tr>
<tr>
<td>Female, n (%)</td>
<td>9 (75.00)</td>
</tr>
<tr>
<td>Age (year)</td>
<td>55±19.64</td>
</tr>
<tr>
<td>Height (cm)</td>
<td>165.58±5.4</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>62.42±9.3</td>
</tr>
<tr>
<td>First surgery age (year)</td>
<td>45.58±20.59</td>
</tr>
<tr>
<td>Implant duration (year)</td>
<td>9.39±4.14</td>
</tr>
<tr>
<td>First surgery reason, n (%)</td>
<td></td>
</tr>
<tr>
<td>Rheumatic heart disease</td>
<td>8 (66.67)</td>
</tr>
<tr>
<td>Infective endocarditis</td>
<td>2 (16.67)</td>
</tr>
<tr>
<td>Degenerative valvular disease</td>
<td>1 (8.33)</td>
</tr>
<tr>
<td>Congenital heart disease</td>
<td>1 (8.33)</td>
</tr>
<tr>
<td>Hypertension, n (%)</td>
<td>5 (41.67)</td>
</tr>
<tr>
<td>Atrial fibrillation, n (%)</td>
<td>4 (33.33)</td>
</tr>
<tr>
<td>Coronary Heart Disease, n (%)</td>
<td>2 (16.67)</td>
</tr>
<tr>
<td>Valve size, n (%)</td>
<td></td>
</tr>
<tr>
<td>25 mm</td>
<td>3 (25.00)</td>
</tr>
<tr>
<td>27 mm</td>
<td>6 (50.00)</td>
</tr>
<tr>
<td>29 mm</td>
<td>3 (25.00)</td>
</tr>
</tbody>
</table>

Values are presented as n (%) or mean ± SD.
Echocardiography demonstrated mitral regurgitation in the porcine mitral valves with cusp tears and commissural dehiscence.

Among the 12 porcine mitral valves, 7 (58.33%) valves showed some degree of calcification. Of these 7 valves, 14.29% (n=1) showed type I calcification, 28.57% (n=2) showed type II calcification, 14.29% (n=1) showed type III calcification and 42.85% (n=3) showed type IV calcification (Figure 3, Table 2). The mean implant duration was 9.94±2.89 years for moderate and severe calcified valves and 9.11±4.8 years for other valves (no significant difference, P=0.763). There was a significant difference in the mean patient age at first implantation between moderate and severe calcified valves and other valves (23.25±18.84 vs. 56.5±9.15 years; P=0.032).

Pannus was found on the leaflet surface in 11 (91.67%)
porcine mitral valves (Figure 4), including 2 valves with type I, 3 with type II, 4 with type III and 2 with type IV (Table 2). In other words, pannus formation on the surfaces of mitral valves appeared to be a universal phenomenon in the explanted porcine mitral valves. However, only one porcine valve was damaged by pannus formation.

**Discussion**

Compared with mechanical valves, the lower risks of thromboembolic and hemorrhagic complications are the major advantage for bioprosthetic valves (14). However, the durability of bioprosthetic valve remains a major factor restricting the wide application in young patients (15,16). Due to the anatomical structure of the mitral valve, the modes of porcine valve deterioration may be affected by left ventricular blood flow and pressure (17-19). Furthermore, due to the asymmetry of porcine native aortic valve leaflets, the structure of porcine valves would affect the stress distribution and blood flow pattern, and consequently affect the long-term durability of porcine valves (20).

Type I cusp tears and commissural dehiscence occurred near the stent post position in 91.67% of explanted valves which was the main mode of failure in the porcine mitral valves in the present study. The damage rate near the stent posts was higher than that reported by Naqvi et al. (21) (91.67% vs. 42%). This may be due to patients who undergo the first surgery at younger age, high prevalence rate of rheumatic heart disease, the structure of bioprosthetic porcine valve, and left ventricular stresses. The patient's average age at first surgery was lower than that reported by Naqvi et al. (21) (45.42±19.58 vs. 58.53±12.4 years). In young patients, energetic physical activity may be an important reason of valve tear. In our study, 66.67% (n=8) patients underwent mitral valve replacement due to rheumatic heart disease which accounts for a major portion of valvular disease in China (22,23). We found that the cusp tear was the only failure reason of explants from rheumatic heart disease patients, while the failure modes of explants from other patients who without rheumatic heart disease were usually tears caused by calcification. The rheumatic disease may affect the strength of collagenous fibers of leaflets, result in cusp tears. Chew et al. (24), who studied...
the mechanical features of porcine valve models, found that the porcine valve was subjected to the highest stress near the stent post area leading to leaflet tears, and that the stress at the tip of the tear increased with the length of the tear so that the degree of cusp tear would increase. Sun et al. (25) took advantage of the finite element method and experimental measurements to confirm that the maximum stress on the tri-leaflet valve was located near the stent posts in the closed state. The localized stress concentration at the stent position occurred in both porcine and bovine pericardial valves. However, the rate of cusp tears was lower among bovine pericardial valves. In a study of bovine pericardial valves, leaflet tears near the stent post area constituted a low proportion of the explanted valves (17.36%) (9). It has been indicated that porcine valves have thinner leaflets and lower mechanical properties than bovine pericardial valves (26). Bovine pericardial valves can bear higher levels of stress due to the material properties near the stent posts; however, porcine valves are more susceptible to leaflet tears. The method of sutureing porcine valve has also been proposed as a factor that contributes to commercial dehiscence. Bottio et al. (10) showed that the mode of stitching the leaflets onto the stent may affect commissural dehiscence in different types of porcine valves. The stress concentration at the sutures also undermines the stability of valve leaflets near the stent posts, leading to commercial dehiscence at the attachment site of the porcine valve to the stent (3). In a study of porcine aortic valves (aortic valves, 61.3%; mitral valves, 35.6%; and others, 3.1%), Butany et al. (27) showed that 71.16% of porcine valves were damaged near the stent posts and the rate of mitral valve rupture was greater than that of aortic valve rupture. The left ventricle exerted greater pressure on the mitral valve than on the aortic valve. When the valve was closed, the orifice area of the mitral valve was larger than that of the aortic valve. Consequently, the total stress on the mitral valve was larger than that on the aortic valve during the entire cardiac cycle. In China, a large proportion of patients suffered from hypertension and the awareness and control rates of hypertension remained lower than in North American (28). In our study, 41.67% of the patients had hypertension. Hypertension increases the stretch ratio of the valve leaflets and causes the leaflets to tear easily.

Calcification was noted in 58.33% of the extracted porcine mitral valves and all patients with first surgery age less than 50 years have severe (type IV) leaflet calcification. Valve calcification was correlated with age at first implantation but not with implant duration in our study. These were also shown in Naqvi et al. study (21). Younger patients with bioprosthetic valves were more prone to valve deterioration induced by calcification. High rate of rheumatic heart disease as the reason of first mitral valve replacement was shown in our study. Rheumatic heart disease with young age accounts for a major portion of valvular disease in China (22,23). With the lack of experience with mitral valve repair technology in the past decade, some young women underwent bioprosthetic valves implantations due to fertility requirements, which also contributed to the younger age of the patients. Although the specific mechanism responsible for this observation is unclear, it could be hypothesized that increased calcium metabolism and serum osteocalcin levels in young adults are responsible for the acceleration in valve calcification (29,30).

In a study of 45 porcine aortic valves, Butany et al. (31) noted that more severe and earlier calcification was observed in aortic valves than in mitral valves (P=0.03). It appears that the porcine mitral valve may be damaged by left ventricular pressure and blood flow in the early period of calcification formation. Hemodynamic disorders may not be observed even in moderate formations of calcification in porcine aortic valves. In severely calcified valves, calcified tissues may drop from the calcified leaflets, leading to embolization (32). In contrast to the X-ray findings, some calcification spots were found in the valve leaflets on micro-CT scans (Figure 3). This may have led to the neglect of small calcified tissues and underestimation of the degree of calcification in previous studies. It is worth studying the classification and evaluation of calcified tissue further.

Pannus is mainly characterized by collagen and elastic fibrous tissue covering the valve surface, which thickens and fuses the valve leaflets, thereby limiting the movement of the valve leaflet and further leading to leaflets stiffness and valve stenosis (7). Pannus was presented in 91.67% of the valves, whereas valve deterioration was noted in only one valve (10%). Butany et al. (27) reported that the probability of pannus formation was 95.7% among porcine valves. In addition, Kuniyoshi et al. indicated that 75.86% of mitral bioprosthetic valves had pannus. Pannus formations were prevalent in bioprosthetic valves which were attached to the fabric of the annulus and valve stent. Although the specific mechanism underlying the formation of pannus remains unclear, it may entail a non-immune inflammatory response to foreign bodies (33). Several studies have confirmed that the following factors contribute to pannus formation: the prosthetic valve design, surgical technique, small annulus, and flow turbulence (34-36).
Study Limitations
The present study investigated porcine mitral valves after explantation for valve deterioration. After valve deterioration, some people may decide not to undergo a second valve replacement operation or die due to other reasons. In addition, the progress of new interventional valve replacement technologies such as transcatheter valve-in-valve implantation has eliminated the need for a second thoracotomy, preventing explantation of the damaged valve. Therefore, it was difficult to obtain enough valves. Furthermore, valve deterioration is a complex process associated with a vast variety of risk factors. These risk factors could not be considered in the present study due to patient-related characteristics.

Conclusions
The present study demonstrated that leaflet damage near the stent posts area was the main failure mode of porcine mitral valves in this study. Patients who undergo the first surgery at younger age, higher prevalence rate of rheumatic heart disease, the structure of bioprosthetic porcine valve, and left ventricular stresses could be considered as the main factors causing valve deterioration could be considered as the main factors causing valve damage near the stent post. Valve calcification was correlated with age at first implantation but not with implant duration in our study. Patients who undergo the first surgery at younger age was due to the high rate of rheumatic heart disease and younger women with fertility requirements. Therefore, we should fully consider the clinical factors and structure features of the mitral bioprosthetic valves in future studies.

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Footnote
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Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/jtd-20-3578). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All patients in this study were provided informed consent for undergoing the procedures. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study design was approved by the Ethics Review Committee of Beijing Anzhen Hospital (No. 2020069x).

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References


